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Four new compounds from the roots of Uvaria macrophylla

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Four new compounds, uvamalols A–C (1–3) and uvarimacrophin A (4), have been isolated from the roots of *Uvaria macrophylla*. Their structures have been elucidated by spectroscopic methods. The relative configurations of uvamalols A–C have been established by NOE experiments, and the relative stereochemistry of uvarimacrophin A inferred from the diagnostic NMR data by comparison with known model compounds.

Keywords: Uvaria macrophylla; Annonaceae; Uvamalols A, B and C; Uvarimacrophin A

1. Introduction

Uvaria macrophylla belongs to the genus *Uvaria* of the family Annonaceae, widely distributed in Hainan, Guangdong, and Guangxi provinces of southern China [1]. Acetogenins, alkaloids, dihydroflavonoids, polyoxygenated cyclohexenes and seco-cyclohexenes have been isolated from the genus *Uvaria* in our previous phytochemical investigations [2–7]. Here we describe the structural elucidation of three new polyoxygenated cyclohexenes, uvamalols A-C (1–3), and a new acetogenin, uvarimacrophin A (4), isolated from the title plant.

2. Results and discussion

Uvamalol A (1) has been isolated as a white powder. Its HR-EIMS gave a protonated molecular ion peak $[M + H]^+$ at m/z 385.1281, which is consistent with the molecular formula $C_{21}H_{20}O_7$.

The IR spectrum for **1** displays absorption bands attributable to hydroxyl (3506 and 3402 cm^{-1}), carbonyl (1714 and 1705 cm⁻¹) groups, and benzene ring (1601 and 1509 cm⁻¹). The ¹H NMR spectrum exhibits the presence of four oxygenated methine protons and a pair of germinal oxygenated methylene protons at δ 3.70–5.20 and an olefinic proton at δ 5.71 (1H, s).

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In combination with the observed signals for two olefinic carbons at δ 136.8 and 128.6, and four oxygenated tertiary carbons at δ 71.0, 73.6, 75.8, and 78.7, as well as a secondary oxygenated carbon at δ 65.1 in the ¹³C NMR spectrum, it is suggested that **1** is a polyoxygenated cyclohexene with a $\Delta^{1(6)}$ double bond [3,5]. In addition, the ten aromatic proton signals at δ 7.42–8.02 (10H, m), 14 carbon signals at δ 168.0, 167.3, 134.4, 134.1, 131.8, 131.3, 130.7 (2C), 130.6 (2C), 129.6 (2C), and 129.4 (2C) in the ¹H and ¹³C NMR spectra, together with a basic peak at *m/z* 105 in the FABMS, indicate the presence of two benzoyl groups [5,7].

In the HMBC spectrum, correlations of the proton signals at δ 4.98 (1H, d, J = 13.5 Hz) and 4.82 (1H, d, J = 13.5 Hz) with the carbonyl signal at δ 167.3 reveal that a benzoyl is attached to 7-OH, while correlations of the proton signals at δ 5.71 (1H, s) and 4.41 (1H, d, J = 8.0 Hz) with the carbon signal at δ 78.7, as well as the proton signal at δ 5.15 (1H, dd, J = 8.0, 11.0 Hz) with the carbonyl signal at δ 168.0, suggest that another benzoyl is linked to 4-OH (figure 1).

The relative stereochemistry of **1** was established by the inspection of NOE correlations and the observed coupling constants. The *J* values equal to or more than 8.0 Hz between H-2, H-3, H-4, and H-5 suggested that they each were all in trans relationship. This was also deduced by NOE difference experiments, the signals at δ 3.72 (H-3) and 5.71 (H-6) showed NOE enhancements when the proton at δ 4.41 (H-5) was irradiated, similarly the signal at δ 5.15 (H-4) also showed NOE enhancement when the proton at δ 4.30 (H-2) was irradiated.

Uvamalol B (2) was obtained as white crystals, its HR-EIMS revealed a quasi-molecular ion peak at m/z [M + H]⁺ 443.1716, corresponding to the molecular formula C₂₄H₂₆O₈.

Absorption bands at 3249, 1730, 1600, 1495 cm⁻¹ in the IR spectrum indicate the presence of hydroxyl, carbonyl, and benzene ring groups. Detailed examination of the ¹H and ¹³C NMR spectra reveal the presence of benzoyl, (2-hydroxyl-5-methoxyl)benzyl, and acetyl functionalities [3,5,6]. The five aromatic protons at δ 7.58–8.01, together with the carbon signals at δ 165.9, 133.3, 129.7, 129.6, and 128.5, are ascribed to a benzoyl. The ABX system proton signals at δ 6.77 (1H, d, J = 8.0 Hz), 6.68 (1H, dd, J = 8.0, 3.0 Hz) and 6.63 (1H, d, J = 3.0 Hz), a methoxyl signal at δ 3.74 (3H, s) and a pair of methylene protons at δ 2.74 (1H, dd, J = 17.0, 7.0) and 3.07(1H, dd, J = 17.0, 7.0), as well as the carbon signals at δ 153.9, 147.4, 122.1, 117.5, 113.8, 113.4, 55.6, and 23.5, are characteristic of a (2-hydroxyl-5methoxyl)benzyl unit [6]. Resonances for the protons at $\delta 2.10$ (3H, s) and carbons at $\delta 20.9$ and 172.0 are assigned to an acetyl. The remaining NMR signals (table 1) are closely comparable to those of polyoxygenated cyclohexenes with a $\Delta^{1(6)}$ double bond [3], in particular the signals for the proton at δ 2.67 (1H, dd, J = 7.0, 4.0) and its corresponding carbon signal at δ 35.2 suggest that C-3 in the cyclohexene unit does not bear oxygen. ${}^{1}H^{-1}H$ COSY correlations of the proton signals at δ 5.57 (H-5) with δ 4.08 (H-4), and δ 2.67 (H-3) with δ 4.08 (H-4), 2.74 (H-8) and 3.07 (H-8), indicate that the benzyl unit should be linked to

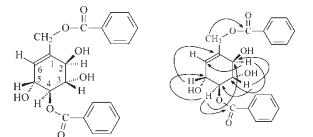


Figure 1. Structure and key HMBC correlations of uvamalol A (1).

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Table 1. ¹H and ¹³C NMR spectral data for 1 in CD₃OD, 2 and 3 in CDCl₃^a.

Position	1		2		3	
	$\delta_H J (Hz)$	δ_C	$\delta_H J (Hz)$	δ_C	$\delta_H J (Hz)$	δ_C
1		136.8		132.3		135.3
2	4.30, d (8.0)	73.6	4.77, s	70.8	4.57, s	67.7
2 3	3.72, dd (10.0, 8.0)	75.8	2.67, dd (7.0, 4.0)	35.2	5.42, dd (6.0, 4.0)	77.0
4	5.15, dd (10.0, 8.0)	78.7	4.08, dd (4.0, 5.5)	73.3	2.86, m	33.5
5	4.41, d (8.0)	71.0	5.57, d (5.5)	72.9	5.01, s	70.2
6	5.71, s	128.6	6.24, s	131.2	6.14, s	129.4
7	4.82, d (13.5)	65.1	4.76, d (12.0)	64.2	4.81, d (13.0)	64.7
7	4.98, d (13.5)		4.93, d (12.0)		5.18, d (13.0)	
8			2.74, dd (17.0, 7.0)	23.5	2.98, dd (16.5, 8.5)	23.6
8			3.07, dd (17.0, 7.0)		2.76, dd (16.5, 8.5)	
9				122.1		120.9
10				147.4		147.3
11			6.77, d (8.0)	117.5	6.77, d (7.0)	117.3
12			6.68, dd (8.0, 3.0)	113.8	6.66, dd (7.0, 2.5)	113.7
13				153.9		153.4
14			6.63, d (3.0)	113.4	6.56, d (2.5)	113.6
1'		168.0		165.9		166.6
1″		134.4		129.6		129.5
2" (6")	8.01, m	130.7	8.01, m	129.7	8.01, m	129.7
3″ (5″)	7.44, m	129.6	7.44, m	128.5	7.44, m	128.3
4″	7.58, m	131.8	7.58, m	133.3	7.57, m	133.3
1‴		167.3				166.6
1////	8.02, m	134.1				129.6
2""(6"")	7.46, m	130.6			8.03, m	129.7
3""(5"")	7.60, m	129.4			7.46, m	128.4
4″″`	·	131.3			7.59, m	133.3
AcO			2.10, s	20.9	·	
			*	172.0		
OCH ₃			3.74, s	55.6	3.75, s	55.7

^a Chemical shift values are in ppm, and J values in parentheses are in Hz. Assignments were confirmed by ${}^{1}H-{}^{1}H$ COSY, HSQC, and HMBC experiments.

C-3 (figure 2). HMBC correlations of the protons at δ 4.76 (H-7) and 4.91 (H-7) with the carbon signal at δ 165.9 (C-1'), and of the proton at δ 5.57 (H-5) with the carbon at δ 172.0, reveal that the benzoyl and acetyl are attached to 7-OH and 5-OH, respectively.

The relative stereochemistry of **2** was interpreted by the NOESY experiment. In the NOESY spectrum, the signal for the proton at δ 5.57 (H-5), correlating with protons at δ 2.74 (H-8), indicates that the benzyl and H-5 are on the same side of cyclohexene and axially

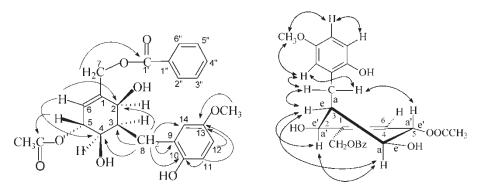


Figure 2. Key HMBC and NOE correlations of uvamalol B (2).

oriented; meanwhile, the signal for the proton at $\delta 2.67$ (H-3) is correlated to the signals for the protons at $\delta 4.77$ (H-2) and $\delta 4.08$ (H-4), suggesting an equatorial bond for H-3, and axial bonds for H-2 and H-4 located on the other side of the cyclohexene.

Uvamalol C (3) was isolated as an oil, and its molecular formula was assigned as $C_{29}H_{28}O_8$ on the basis of its HR-FABMS ($[M - H_2O]^+ m/z$ 486.1678).

A preliminary examination of the spectral data of **3** suggested that this compound is very similar to **2**, both having a polyoxygenated cyclohexene moiety, a (2-hydroxy-5-methoxyl)benzyl unit and a benzoyl group [3,5–7]. However, there are some significant differences between the two compounds: **3** has one benzoyl more than **2** and no acetyl group, moreover, the chemical shifts of the cyclohexene proton signals in **3** are apparently different from those in **2**, indicating that the benzyl unit is not substituted to C-3. In the ${}^{1}\text{H}-{}^{1}\text{H}$ COSY spectrum, the proton signal at δ 2.86 (H-4) is correlated to the protons at δ 5.42 (H-3), 2.98 (H-8), and 2.76 (H-8). The HMBC experiment clearly exhibited long-range correlations of the protons at δ 2.98 (H-8) and 2.76 (H-8) with the carbons at δ 77.0 (C-3), 33.5 (C-4), and 70.2 (C-5), and the proton at δ 6.14 (H-6) with the carbons at δ 70.2(C-5) and 33.5(C-4) (figure 3). These findings reveal that the benzyl unit is linked to C-4.

In the NOE spectrum, significant cross peaks of H-4 with H-5 and H-3 suggest an equatorial bond for H-4 and axial bonds for H-3 and H-5 on the same side of the hexene ring; a cross peak of H-2 with H-8 reveals that the benzyl unit and H-2 are both axially oriented on the other side of the hexene ring.

Uvarimacrophin A (4) was isolated as a waxy solid, and its molecular formula was determined as $C_{39}H_{70}O_7$ by HR-FABMS ([M + H]⁺ m/z 651.5179).

A prominent IR carbonyl absorption at 1755 cm⁻¹ suggests the presence of an unsaturated γ lactone, which is confirmed from the NMR spectra, showing diagnostic signals at δ 6.95 (1H, H-35), 4.99 (1H, H-36), 1.41 (3H, H-37) and δ 173.8 (C-1), 148.8 (C-35), 134.3 (C-2), 77.3 (C-36), 19.2 (C-37) [4,8]. Both the IR carbonyl absorption band at 1720 cm⁻¹ and the resonance for the protons at δ 2.06 (3H, s) and the carbons at δ 171.5, and 21.2 reveal the presence of an acetoxyl group, which is also supported by the fragment ion at m/z 591 ([MH – HOAc]⁺) in the FABMS spectrum [4,8]. Proton signals at δ 5.11 (1H, H-17), 3.86 (1H, H-18), 3.82 (1H, H-21), and 3.42 (1H, H-22), and the corresponding carbon signals at δ 71.6 (C-17), 82.2 (C-18), 82.9 (C-21), and 74.1 (C-22), are characteristic for mono-THF acetogenin with one flanking hydroxyl and another flanking acetoxyl [4,8]. In addition, successive losses of two H₂O units from the fragment ([MH – HOAc]⁺) in the FABMS indicates the presence of another hydroxyl group.

The locations of the THF ring, the two hydroxyls and the acetoxyl along the aliphatic chain in **4** were determined by analysis of the EIMS of **4** and **4a** (figure 4). EIMS of **4/4a** gave fragment

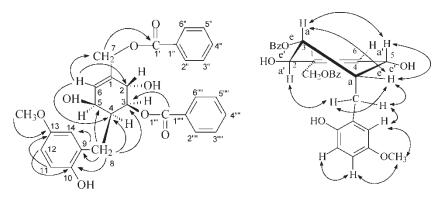
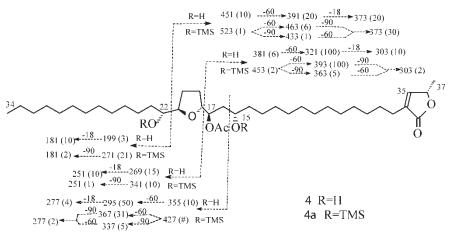


Figure 3. Key HMBC and NOE correlations of uvamalol C (3).

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ion peak not observed

Figure 4. Structure of uvarimacrophin A (4) and diagnostic EIMS fragment ions (m/z) of 4 and 4a.

ions at m/z 295/367, 381/453, and 453/523, which clearly show the position of the THF ring between C-18 and C-21, two hydroxyls at C-15 and C-22, and the acetoxyl at C-17 [4]. Correlation of the proton at δ 3.38 (H-15) with that at δ 5.11 (H-17) through H-16 (δ 1.54) in the ¹H-¹H COSY spectrum substantiates the location of the isolated hydroxyl at C-15; meanwhile, a significant cross peak between the proton at δ 5.11(H-17) and the carbonyl carbon at δ 171.5 in an HMBC experiment also demonstrates that the acetoxyl is linked to C-17.

Based on the foregoing discussion, the diagnostic NMR data is closely similar to those of calaminstrin B, except for the signals of H-22 and C-22, indicating that the relative stereochemistries at C-15, C-17, C-18, and C-21 were the same as those of the corresponding carbons in calaminstrin B [4]. Further comparison with diagnostic chemical shifts of a pair of model mono-THF compounds with adjacent hydroxyls in *threo* and *erythro* configurations suggested that the relative stereochemistry between C-21 and C-22 was *threo*, which is evident from the characteristic NMR signals for H-21 at δ 3.82 and C-22 at δ 74.1 [8,9].

The results of the present investigation suggest that uvamalols B and C are a new type of polyoxygenated hexene; its skeleton includes two moieties, a seven carbon cyclohexene unit and a seven carbon benzyl unit, which are linked through a carbon–carbon bond. Interestingly, in combination with our previous investigation, macrophyllols A and B also contain the (2-hydroxy-5-methoxyl)benzyl unit [2,6] — how is the same structural unit attached to two different types of compounds? It is worth exploring the biogenetic pathway of these compounds.

3. Experimental

3.1 General experimental procedures

Melting points were determined on an XT-4 micro-melting point apparatus and are uncorrected. Optical rotations were measured on a Perkin–Elmer 241 polarimeter. IR spectra were recorded on a Perkin–Elmer 683 FT infrared spectrometer. UV spectra were obtained on a Shimadzu UV-240 instrument. NMR spectra were run on a Varian INOVA-500 NMR spectrometer with TMS as internal standard. EIMS were obtained on a VG ZAB-2F mass

spectrometer and ESIMS, FABMS, HR-EIMS and HR-FABMS were performed on an Autospec-Utima ETOF Spec mass spectrometer. HPLC separations were conducted on a TSP 3200 pump system equipped with a TSP 3200 UV detector and with a Waters μ -Bondapak C-18 Hyperprep. Column: 10 μ m, 25 mm i.d. 250 mm long. Compounds were detected by UV radiation at 254 nm.

3.2 Plant material

Roots of *Uvaria macrophylla* were collected in Jianfengling, Hainan Province, China, in December 2000 and identified by Professor Shi-Man Huang. A voucher specimen (No. 62161) has been deposited in the Herbarium of Institute of Materia Medica, Chinese Academy of Medical Sciences.

3.3 Extraction and isolation

Dried, ground roots (10.0 kg) were extracted with 95% EtOH to afford 1.4 kg of a residue on removal of solvent under reduced pressure. The EtOH extract was partitioned between H_2O and CHCl₃, giving a water-soluble fraction I (294 g) and a chloroform-soluble fraction II (230 g) as well as an insoluble fraction III (800 g). Fraction II was first dissolved in 90% MeOH and then defatted with light petroleum to give a methanol-soluble fraction IV (175 g). Fraction IV was subjected to column chromatography on silica gel, eluting with a gradient of light petroleum-acetone, and the eluate collected in 1000 mL fractions (fractions 1-110). Fractions 36-42 + 68-74 and 43-53 showed the presence of polyoxygenated cyclohexenes and acetogenins, respectively, as detected by TLC with a 10% H₂SO₄ alcohol solution containing 0.5% vanillin as spray agent. Fractions 36-42 and 68-74 were mixed and evaporated *in vacuo*, yielding 15.2 g of a residue. The residue was further subjected to column chromatography on silica gel (600g) eluted with light petroleum-EtOAc $(6:4 \rightarrow 3:7)$ in 250 mL fractions (fractions 1–36). Eluted subfractions 9 and 23 contained compounds 2 and 1 + 3, respectively. Subfraction 9 was further purified by preparative TLC with light petroleum–CHCl₃–EtOAc (3:1:2) to give 6.8 mg of $2 (R_f = 0.61)$. Subfraction 23 (8.0 mg) showed a single spot on normal phase TLC, but the NMR spectrum revealed a mixture of two compounds in a ratio of 2:1, therefore this fraction was subjected to semipreparative reversed-phase HPLC (flow rate = 1.0 mL min^{-1}) with MeOH-H₂O (75:25) to afford 5.0 mg of 1 ($t_{\rm R} = 5.37$ min) and 1.5 mg of 3 ($t_{\rm R} = 6.35$ min). The combined fractions 43-53, containing acetogenins, was repeatedly chromatographed on a silica gel column, alternately eluted with light petroleum-Me₂CO (8:1) and light petroleum-EtOAc (5:1), to yield 15.0 mg of **4**.

3.3.1 Uvamalol A. White powder, mp 136–138°C; $[\alpha]_D^{23} - 78 (c0.24, MeOH)$; IR (KBr) ν_{max} (cm⁻¹): 3506 (OH), 3402 (OH), 1714 (C=O), 1705 (C=O), 1601, 1509, 1450, 1315, 1279, 1120, 1070, 1007, 960, 866, 708; UV (MeOH) λ_{max} (nm) (log ϵ): 230 (4.12), 205 (3.04); ¹H (CDCl₃, 500 MHz) and ¹³C (CDCl₃, 125 MHz) NMR, see table 1; FABMS *m/z*: 385 (16), 367 (52), 179 (18), 167 (15), 154 (30), 149 (5), 136 (34), 105 (100), 95 (95), 69 (51), 55 (70); HR-FABMS *m/z*: 385.1281 [M + H]⁺ (calcd for C₂₁H₂₁O₇, 385.1287).

3.3.2 Uvamalol B. White crystals, mp 150–151°C; $[\alpha]_D^{23}$ – 275 (*c*0.05, MeOH); IR(KBr) ν_{max} (cm⁻¹): 3429 (OH), 3062, 2935, 2881, 1730 (C=O), 1712, 1600, 1495, 1431, 1377,

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1327, 1277, 1250, 1205, 1120, 1045, 1032, 945, 926, 858, 816, 712; UV (MeOH) λ_{max} (nm) (log ϵ): 382 (2.82), 287 (3.76), 230 (4.32); ¹H (CDCl₃, 500 Hz) and ¹³C (CDCl₃, 125 MHz) NMR, see table 1; EIMS *m/z* 424 (40), 364 (1), 334 (2), 302 (3), 260 (5), 242 (100), 211 (2), 181 (2), 136 (43), 105 (90), 91 (2), 77 (42), 51 (12); HR-FABMS *m/z*: 443.1716 [M + H]⁺ (calcd for C₂₄H₂₆O₈, 443.1706).

3.3.3 Uvamalol C. Oil, mp 150–151°C; $[\alpha]_D^{23} - 97$ (*c*0.07, MeOH); IR (film) ν_{max} (cm⁻¹): 3469 (OH), 3062, 2924, 2854, 1716 (C=O), 1601, 1452, 1369, 1315, 1269, 1176, 1070, 1026, 806, 710, 687; UV (MeOH) λ_{max} (nm) (log ϵ): 203 (4.16), 228 (4.04), 290 (3.87), 358 (2.95); ¹H (CDCl₃, 500 MHz) and ¹³C (CDCl₃, 125 MHz) NMR, see table 1; FABMS *m/z*: 486 ([M - H₂O]⁺, 1), 159 (5), 147 (30), 123 (25), 105 (42), 81 (65), 69 (87), 55 (100); HR-FABMS *m/z*: 486.1643 [M - H₂O]⁺ (calcd for C₂₉H₂₆O₇, 486.1678).

3.3.4 Uvarimacrophin A. Wax, $[\alpha]_D^{23} + 31$ (*c* 0.08, MeOH); IR(film) ν_{max} (cm⁻¹): 3469 (OH), 2924, 1852, 1757 (C=O), 1739 (C=O), 1466, 1373, 1317, 1244, 1177, 1074, 1026, 955, 879, 719; ¹H (CDCl₃, 500 MHz) δ (ppm): 2.27 (H-3, t, *J* = 6.8 Hz), 1.10–1.60 (H-4-14 and H-23-33, m), 3.38 (H-15, m), 1.55 (H-16, m), 5.11 (H-17, m), 3.86 (H-18, m), 1.92–1.64 (H-19-20, m), 3.82 (H-21, m), 3.42 (H-22, m), 0.87 (H-34, t, *J* = 6.8 Hz), 6.95 (H-35, d, *J* = 1.5 Hz), 4.99 (H-36, dq, *J* = 1.5, 6.8 Hz), 1.41 (H-37, d, *J* = 6.8 Hz), 2.06 (CH₃CO, s); ¹³C NMR (CDCl₃, 125 MHz) δ (ppm): 173.8 (C-1), 134.3 (C-2), 25.1 (C-3), 22.6–38.7 (C-4-14, 19-20, and 23-33), 70.0 (C-15), 25.3 (C-16), 71.6 (C-17), 82.2 (C-18), 82.9 (C-21), 74.1 (C-22), 14.1 (C-34), 148.8 (C-35), 77.3 (C-36), 19.2 (C-37), 171.5 (CH₃CO), 21.2 (CH₃CO); FABMS *m/z*: 651 ([M + H]⁺, 70), 591 ([M + H - HOAc]⁺, 92), 573 ([M + H - HOAc - H₂O]⁺, 80), 555 ([M + H - HOAc - 2H₂O]⁺, 100), 537 (31), 451 (12), 387 (4), 374 (10), 321 (20), 295 (20), 277 (3), 239 (10), 209 (4), 161 (3), 149 (15), 109 (50), 95 (72), 81 (80), 69 (100); EIMS data see figure 4. HR-FABMS *m/z*: 651.5179 [M + H]⁺ (calcd for C₃₉H₇₁O₇).

3.3.5 TMSi Derivative (4a). A small amount (0.5 mg) of **4** was treated with *N*,*O*-bis(trimethylsilyl)-acetamide-pyridine and heated at 70°C for 30 min to yield a tri-TMSi derivative (**4a**). The mixture was used directly to measure EIMS; for EIMS fragments of **4a**, see figure 4.

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